

Molecular Influenza Surveillance with the FluSurver in GISAID



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1. Background

Interest in new outbreaks as well as regular surveillance of circulating seasonal strains produce a constant flow of influenza sequences that need to be analyzed and interpreted for epidemiological and phenotypic features. Several steps in typical influenza sequence analysis can be automated and we have been actively developing the free online analysis pipeline FluSurver over the last 4 years to facilitate identification and interpretation of mutations in influenza sequences.

2. Materials and Methods

The FluSurver is accessible as webserver (<http://flusurver.bii.a-star.edu.sg/>) or directly from GISAID (beta) where users can upload their sequences for fully automated analysis which includes ultra-fast database searches with TACHYON, alignment with MAFFT, structural modeling with MODELLER, structure viewing with Jmol and several PERL scripts to link mutations to in-house derived databases. These include geographic and temporal frequency of occurrence as well as co-occurrence of mutations, curated literature annotations for >250 known mutation effects such as drug resistance, host receptor specificity, virulence, antigenic drift and antibody escape mutants. We also show the position of the mutation(s) in structural models and highlight if mutations are close to common drug, host receptor or antibody binding sites or if a glycosylation motif is lost or created through a mutation. Notable recent additions/changes are: updated reference strains, ultra-fast database searches using TACHYON, added city-level detail to map view, passage history bias information for selected mutations, integration into GISAID (beta), expansion of help and tutorial sections for guidance of using the results in publications and to avoid over-interpretation.

3. Results

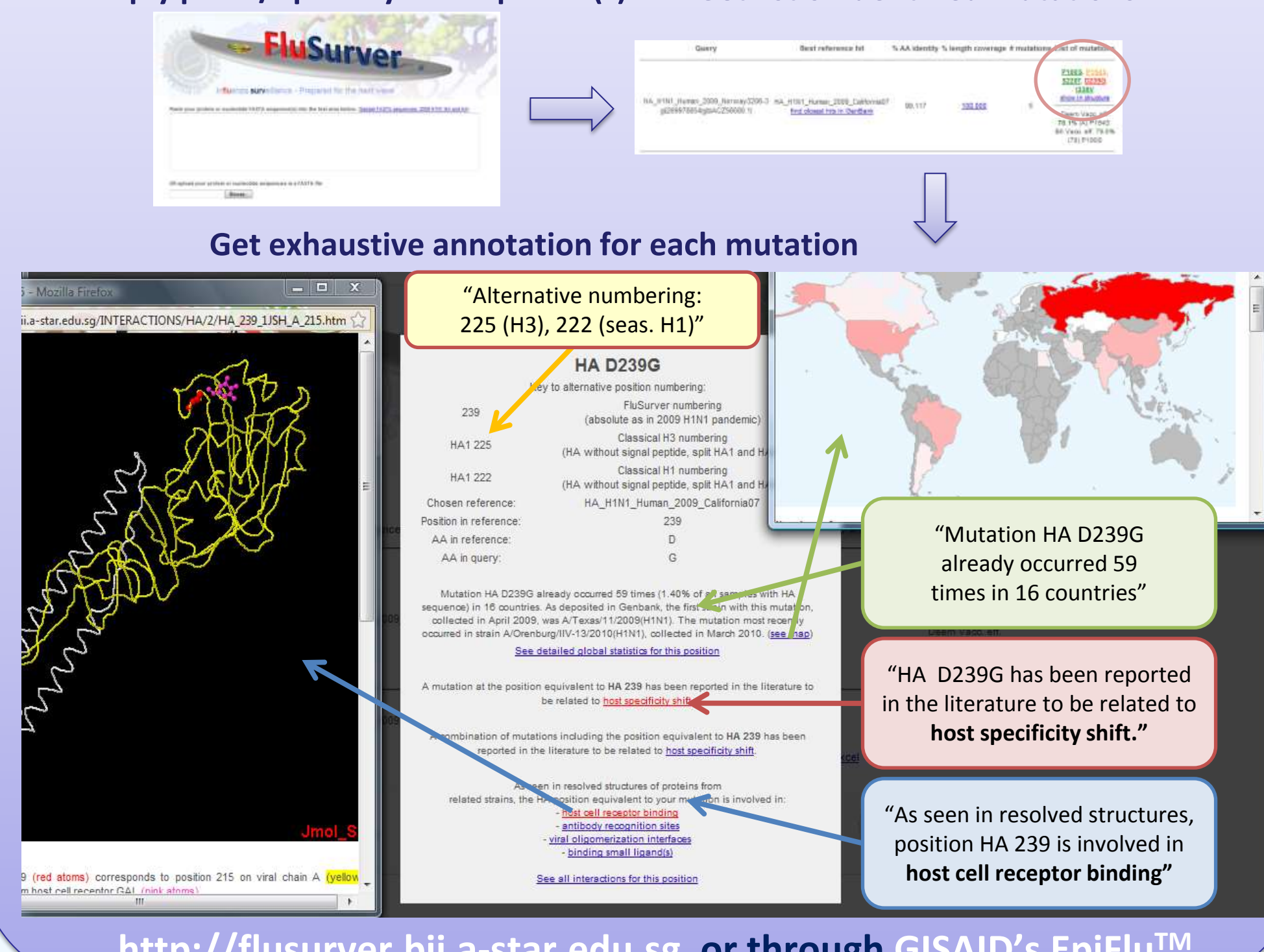
The FluSurver has already been instrumental in the discovery of new influenza strain variants with altered antiviral susceptibility, host specificity, glycosylation and antigenic properties. To showcase the usefulness and speed of analysis made possible with our tool, we use the recent H7N9 outbreak sequences deposited in GISAID as new case study. After submitting the sequences of the first 3 human cases, it only takes a few seconds to identify the critical HA-Q226L and PB2-E627K mutations that are known adaptations facilitating infection of mammalian hosts. Further browsing the alignment results identifies absence of a multi-basic HA cleavage site suggesting low pathogenicity in birds, a small stalk deletion in NA of unclear significance, common resistance to amantadines through M2-S31N, normal neuraminidase drug-sensitivity at NA-H274, one isolated strain with NA-R292K which is associated with drug-treatment induced resistance in closely related avian N9 strains. Another common feature of the early H7N9 outbreak sequences is a truncation of NS1 through an early stop codon resulting in removal of the PDZ-binding motif which is normally used for binding and interfering with host proteins. When including the avian and environmental isolates in the analysis, one can see that while the closest avian precursor strains from 2011-2012 did not have the Q226L host specificity mutation, surprisingly the recent outbreak-linked avian samples already had this mutation which must therefore also be fit to circulate in birds to some extent. At the same time, PB2-E627K was only found in human samples suggesting a stepwise acquisition of host adaptation factors. The FluSurver analysis also highlights unusually diverse strains, e.g. A/Shanghai/1/2013 compared to A/Anhui/1/2013 and A/Shanghai/2/2013, which would indicate multiple introduction events from birds to humans instead of sustained human to human transmission in the beginning of the outbreak.

4. Conclusions

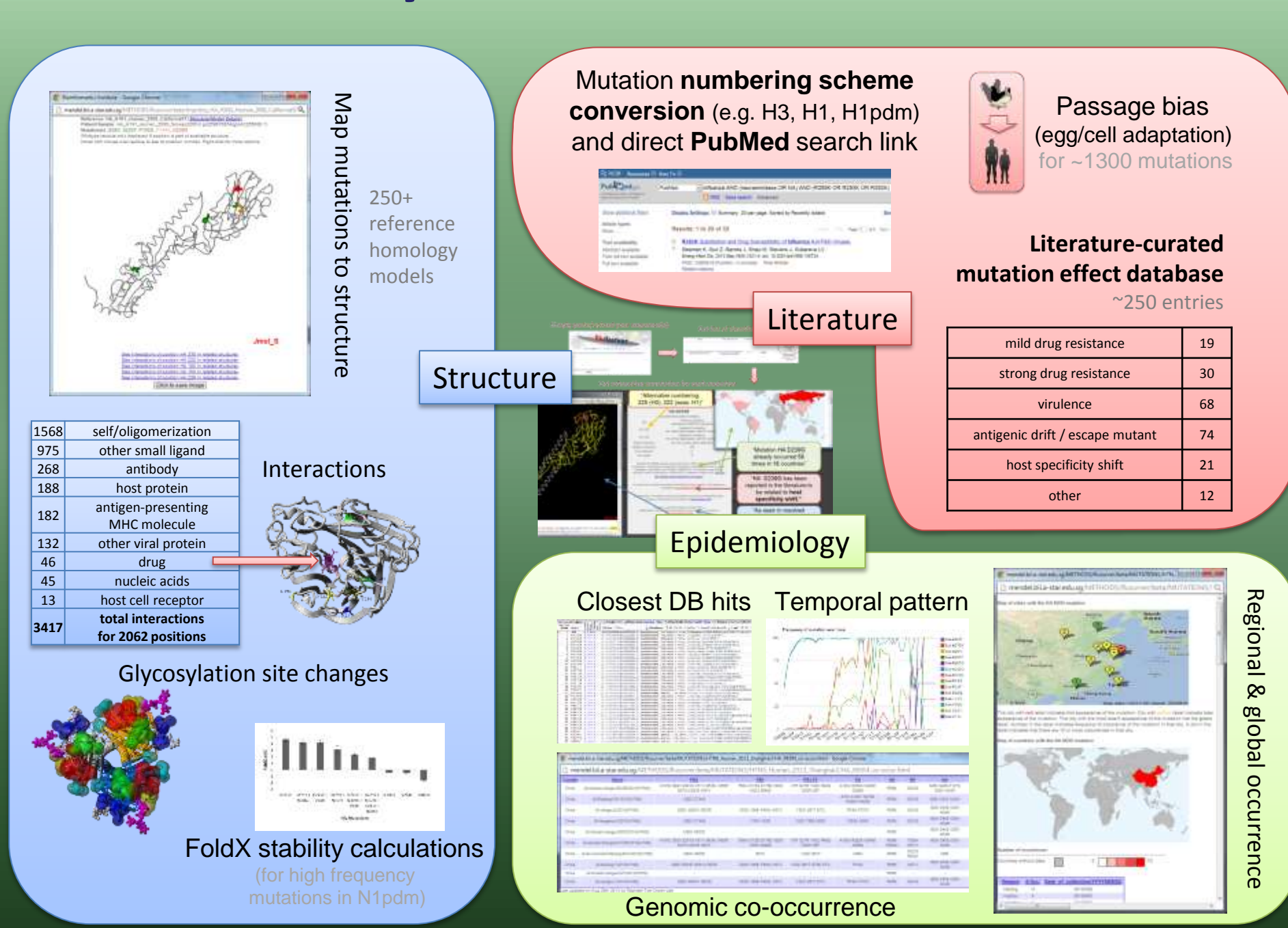
The FluSurver allows researchers, clinician scientists and surveillance labs to rapidly screen their influenza sequences for potentially interesting mutations to identify candidates for phenotypic changes or special epidemiological relevance. In the case of H7N9, it can be used for a solid initial characterization as well as to continue monitoring if additional human host adaptation mutations would occur in the future. The full potential of FluSurver can be realized through the integration with the GISAID database.

FluSurver Basics

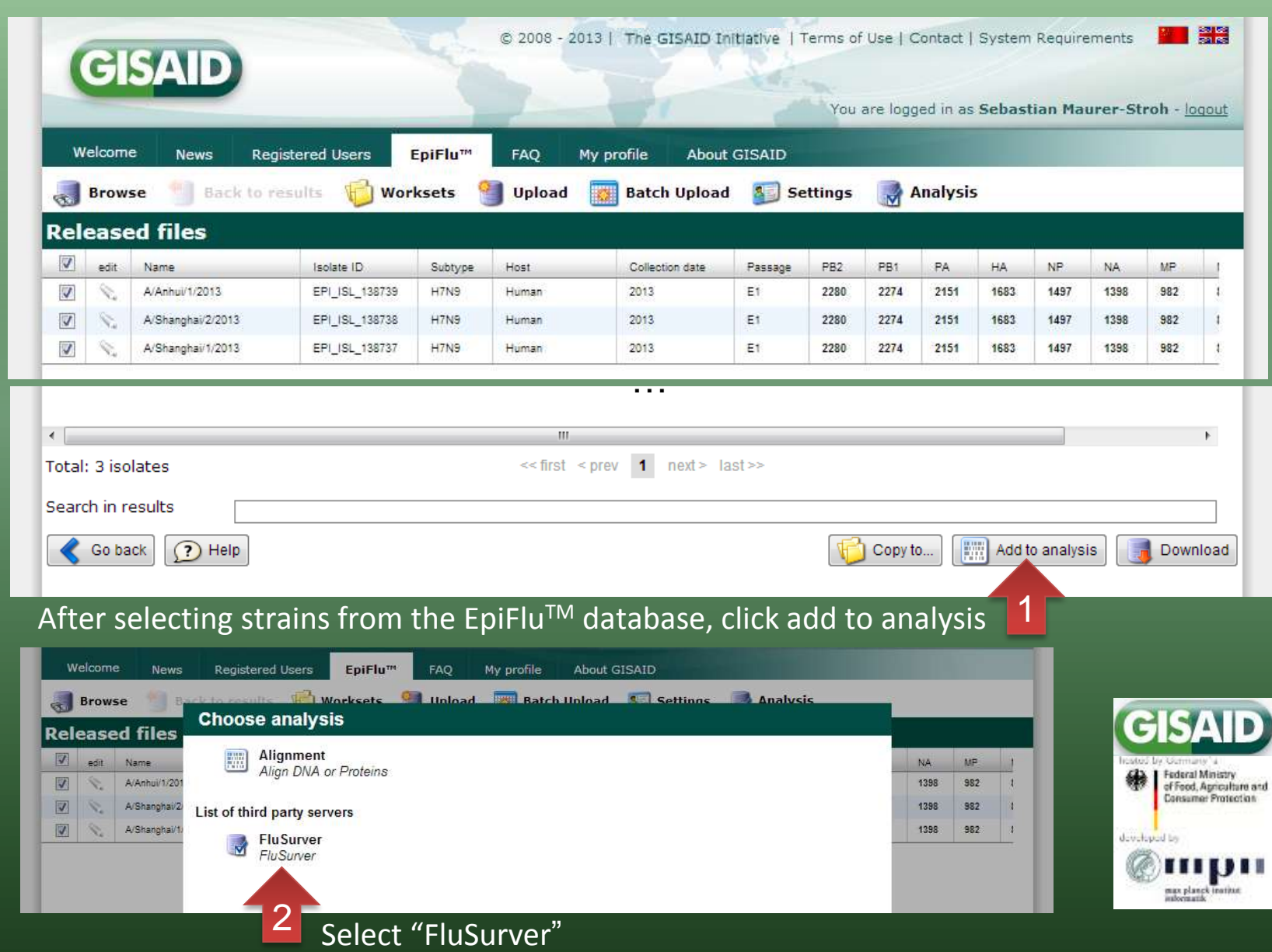
Simply paste/upload your sequence(s): Get list of identified mutations



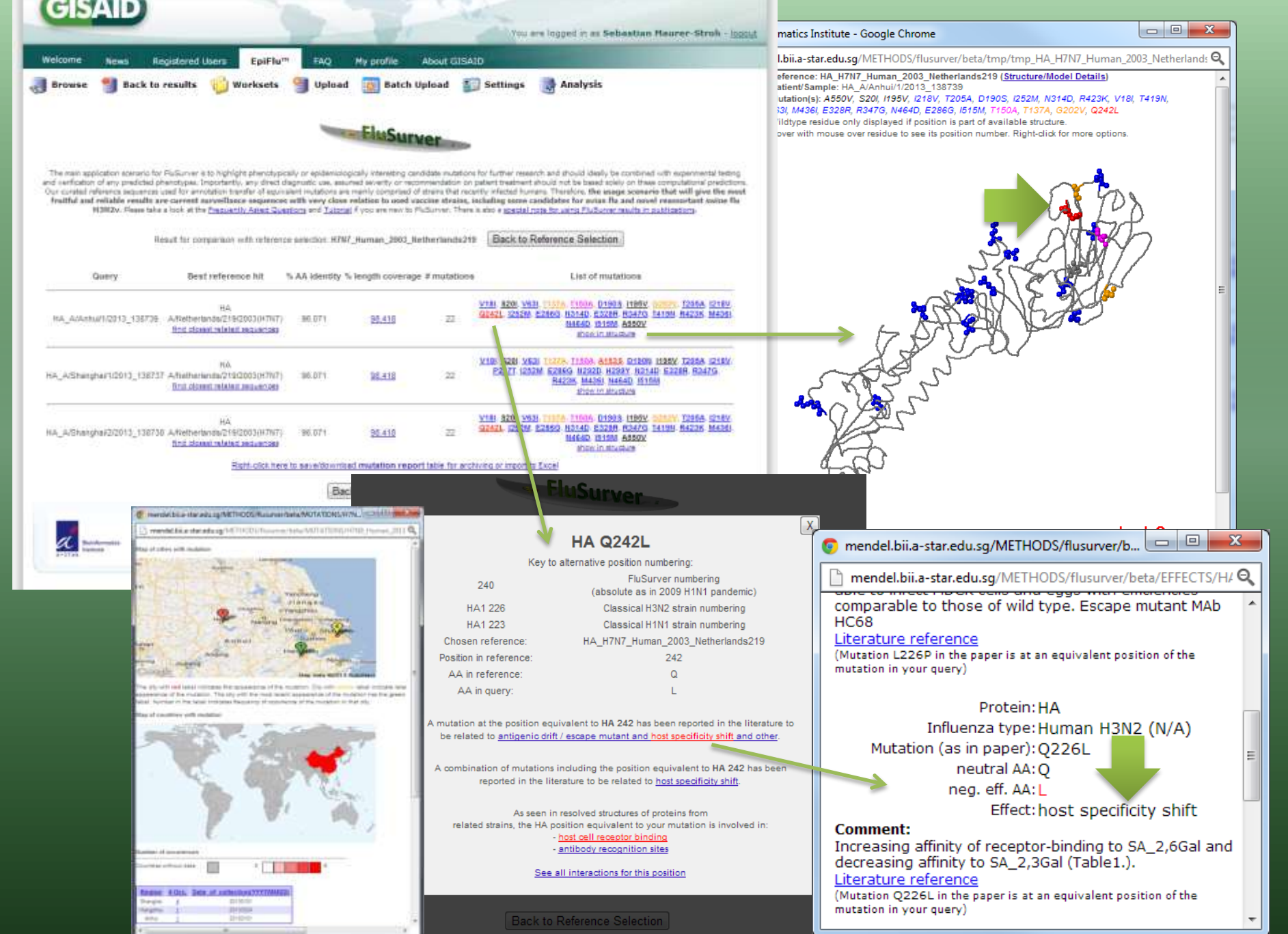
Summary of FluSurver features 2013



NEW (beta test): Analyze sequences with FluSurver directly from the GISAID platform



H7N9 mutation analysis example with FluSurver in GISAID



First 3 H7N9 strains as seen in FluSurver: Summary of major host specificity mutations

Protein-Position(s)	Anhui/1	Shanghai/2	Shanghai/1	Role
HA-226	L	L	Q	Receptor-binding
HA-186	V	V	G	Receptor-binding
HA-138	A	A	S	Receptor-binding
PB2-627	K	K	K	Replication-efficiency

- H7N9 is presumably a **low pathogenic strain in birds** as all 3 HAs lack a multibasic cleavage site

Low patho (e.g. H7N9)
Single protease cleavage site, localized infection

High patho (e.g. H7N7)
Multi protease cleavage site, systemic infection

Segment	Segment	Country	Collection date	Isolate name	Submitting Lab
HA	HA	China	2013-Feb-26	A/Shanghai/1/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Feb-26	A/Shanghai/2/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Feb-26	A/Shanghai/1/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/2/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/1/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/2/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/1/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/2/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/1/2013	WHO Chinese National Influenza Center
HA	HA	China	2013-Mar-05	A/Shanghai/2/2013	WHO Chinese National Influenza Center

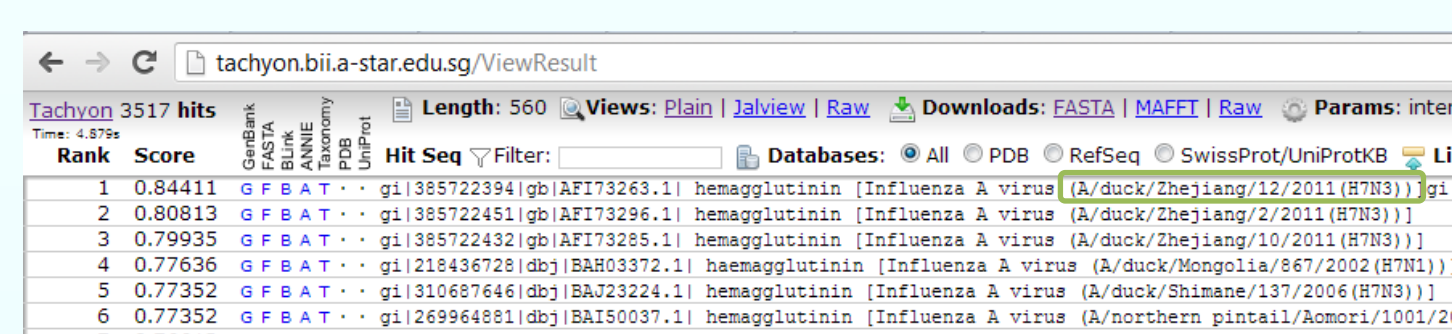
First 3 H7N9 strains as seen in FluSurver: Summary of critical drug sensitivity positions

Protein-Position(s)	Anhui/1	Shanghai/2	Shanghai/1	Role
NA-274	H	H	H	Drug sensitivity (Tamiflu)
NA-292	R	R	K	Drug sensitivity (Tamiflu and Relenza)
NA-58-60	del58-60	del58-60	del58-60	Stalk deletion
M2-31	N	N	N	Drug sensitivity (Amantadine)

Geographic distribution of strains with identified mutations:



Find closest database hits to trace back reassortment events

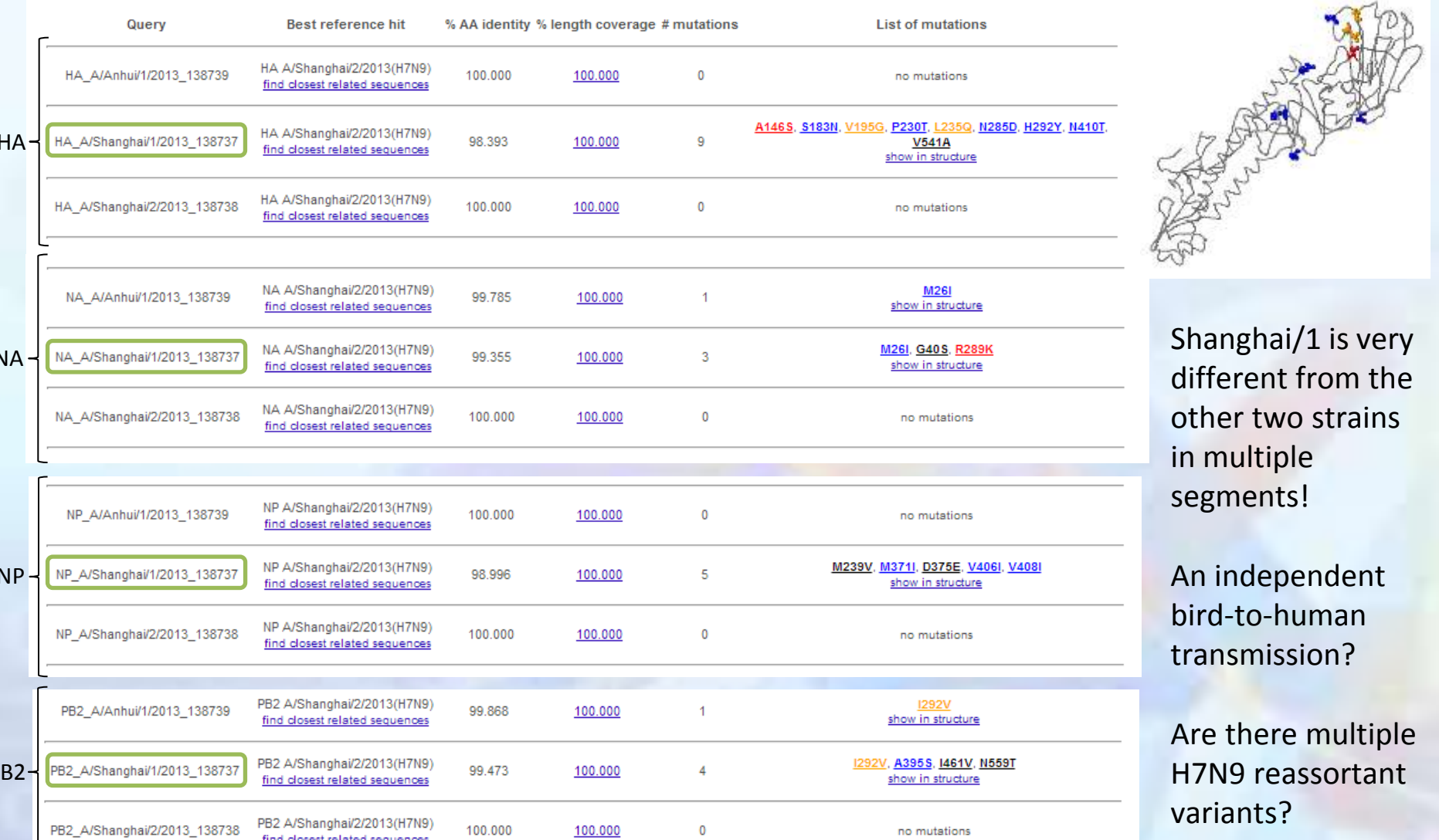


Tachyon is an ultra-fast protein search engine against large databases and can be accessed from inside FluSurver by clicking on “find closest related sequences”.

The consensus of best database hits shows the likely origin of the individual segments:
HA = H7N3
NA = N9...H11N9/H7N9/H10N9/H9N9?
All other = H9N2

BII's Tachyon takes a few seconds while Bayesian phylogenetic trees take hours to days...

Monitor any new mutations or deviating strains in the outbreak

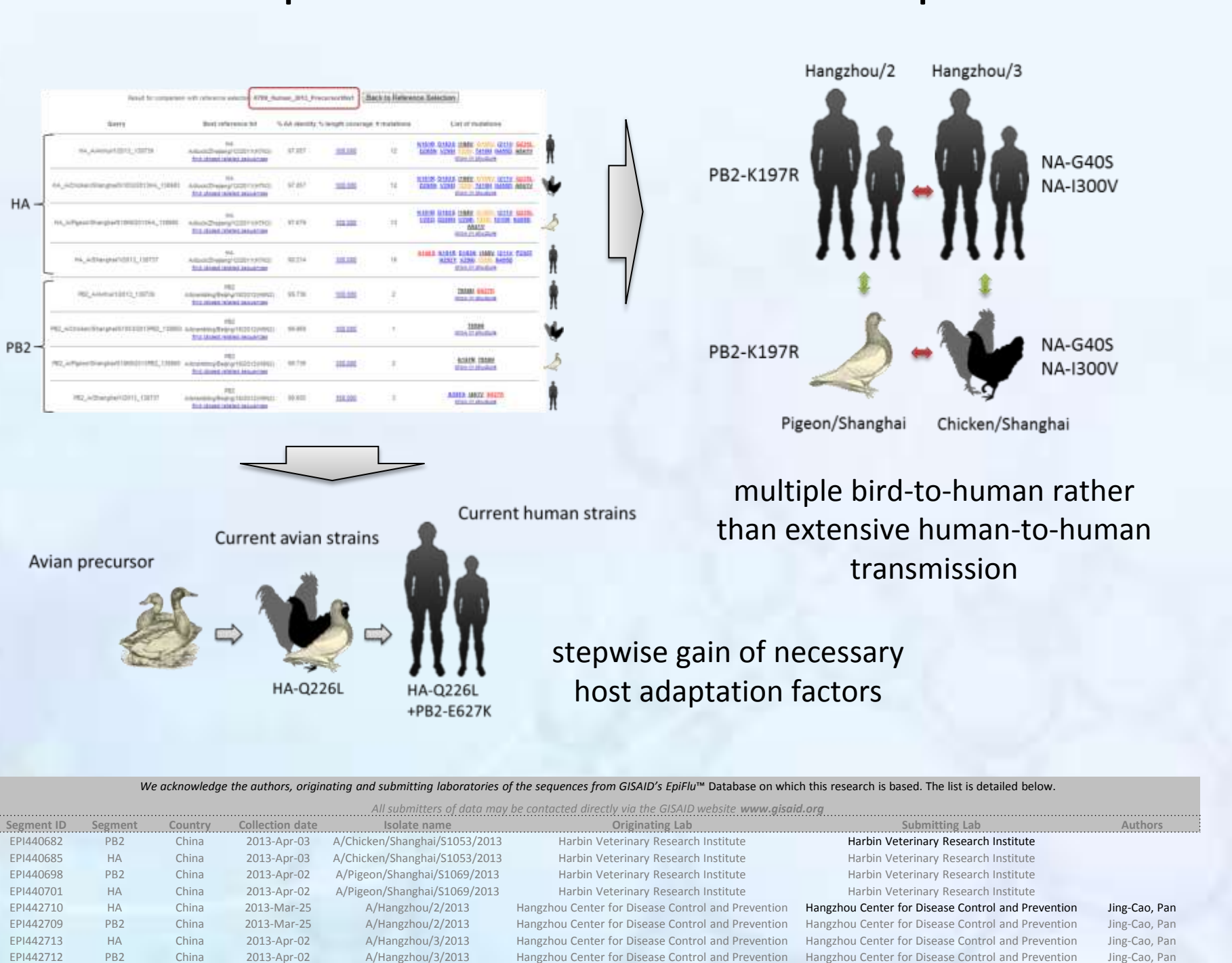


Shanghai/1 is very different from the other two strains in multiple segments!

An independent bird-to-human transmission?

Are there multiple H7N9 reassortant variants?

Compare avian and human samples



Collaborative influenza research where our FluSurver has already been useful:

- Guarnaccia T, Carlson LA, Maurer-Stroh S, Lee RT, Job E, Reading PC, Peters B, McCaw JM, McVernon J, Hurt AC, Kelso A, Mossa J, Barr IG, Laurie KL, Antigenic Drift of the Pandemic 2009 A/H1N1 Influenza Virus in a Ferret Model. PLoS Pathog. 2013 May 9;9(5):e1003254. doi: 10.1371/journal.ppat.1003254. Epub 2013 May 9. PubMed PMID: 23671416.
- Maurer-Stroh S, Lee RT, Gnanan V, Eisenhaber F. The highly pathogenic H7N3 avian influenza strain from July 2012 in Mexico acquired an extended cleavage site upon reassortment with host 285 H3NA. Virol J. 2013 May 1;10:139. doi: 10.1186/1743-4229-10-139. PubMed PMID: 23635055.
- Job ER, Deng YM, Barford K, Tate MD, Colwell N, Redeker S, Maurer-Stroh S, Barr IG, Reading PC. Addition of Glycosylation to Influenza A Virus Hemagglutinin Modulates Antibody-Mediated Recognition of H1N1 2009 Pandemic Virus. J Immunol. 2013 Mar 1;190(3):2109-17. doi: 10.4049/jimmunol.1202433. Epub 2013 Jan 30. PubMed PMID: 23360855.
- Simoes EA, Patel C, Sung WK, Lee CW, Loh KH, Lucero M, Nohynek H, Nair G, Thien PL, Koh CW, Chen YS, Ma J, Maurer-Stroh S, Grassano-Link P, Hibberd ML, Wong CW, IAVIC Consortium. Pathogen chip for respiratory tract infections. J Clin Microbiol. 2013 Mar 5;151(3):845-51. doi: 10.1128/JCM.02371-12. Epub 2012 Jul 20;2012:148-57. Epub 2012 May 4. PubMed PMID: 22561367.
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